

# **Product Datasheet**

# Green Live/Dead Stain (orb511139)

Catalog Number orb511139

**Category** Tools

Description

Green Live/Dead Stain is a vital dye that exhibits intact cell membrane exclusion properties analogous to the popular red fluorescing vital dyes, Propidium Iodide (PI), 7-amino- actinomycin D (7-AAD) and DRAQ7. Like the red fluorescing dyes, Green Live/Dead Stain is excluded from intact, healthy cells due to its polar nature. In the presence of cells exhibiting compromised membrane integrity, Green Live/Dead Stain penetrates the cell and nuclear membrane barriers and intercalates tightly to DNA in a manner analogous to PI and 7-AAD. When bound to DNA, it acquires a greatly enhanced fluorescence potential (>2000X) in the green emission range. These important vital dye properties enable Green Live/Dead Stain to be used in flow cytometry-based protocols to assess the percentage of late apoptotic, necrotic, and membrane-compromised cells within a sample cell population. When bound to nucleic acids, the maximum absorption of Green Live/Dead Stain is 495 nm and the maximum emission is 512 nm. Cells can be viewed through a fluorescence microscope or analyzed with a flow cytometer. Green Live/Dead Stain is provided as a 500 µM concentrated stock solution dissolved in DMSO. For flow cytometry applications, a staining concentration of 50 nM is recommended. Therefore, using sample sizes of 0.5 mL, a single 50 µL vial provides enough reagent for 1000 tests. For fluorescence microscopy applications, a usage concentration of 0.5 μM is suggested. In this way, a vial is sufficient for 100 tests (0.45 mL sample sizes). Green Live/ Dead Stain can be used with red FLICA 660 caspase inhibitor reagents to identify four populations of cells: living; early apoptotic; late apoptotic; and necrotic.

**Target** Dead cells, necrosis

**Concentration** 500 μM

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### **Application notes**

1. Allow product to equilibrate to room temperature (25°C) prior to use., 2. Add stop solution to plate well. Equal volumes of TMB microwell substrate and stop solution should be used. We recommend using 100  $\mu$ L of TMB substrate and 100  $\mu$ L of Stop Solution for TMB Substrates per well. The stopped TMB reaction product is stable for 1 hour., 3. Read absorbance at 450 nm within 60 minutes. For best results, sample absorbance should be monitored and stopped before values exceed 2.0 OD units.

Sample Types Cell culture

Storage -20°C

**Note** For research use only

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