



# Product Datasheet HRP Western Blot Mouse IgG antibody

(Peroxidase) (orb345943)

Description	HRP Western Blot Mouse IgG antibody (Peroxidase)
Species/Host	Goat
Conjugation	HRP
Tested Applications	ELISA, IHC, WB
Preservatives	0.02 M Potassium Phosphate, 0.15 M Sodium Chloride, pH 7.2
Concentration	1.0 mg/ml
Storage	See kit insert for complete instructions.
Note	For research use only
Application notes	Western Blot Analysis
Clonality	Polyclonal
Dilution Range	ELISA: 1:10,000-1:100,000, IHC: 1:500-1:5,000, WB: 1:5,000-1:40,000
Expiration Date	12 months from date of receipt.



(A) LoVo cells were either exposed to PTH $\alpha$  (30 µm) or transfected with non-specific siRNA (ns-siRNA) or p14ARF specific siRNA. Cells were treated with 2.5 µm oxaliplatin 8 h after siRNA or 1 h after PTH $\alpha$  treatment. 120 h upon oxaliplatin exposure, the expression of p14ARF, p21CIP1, and p53, as well as the phosphorylation of p53 at Ser15 was measured by immunodetection. HRP conjugated goat anti-mouse (p/n orb345943) and HRP conjugated goat anti-rabbit (p/n orb345944) were used.

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Longitudinal dynamics of neutralizing and anti-N antibody responses to SARS-CoV-2 infection from outpatient and hospitalized individuals. a, b. The half-maximum inhibitory concentration (IC50) of sera was determined by microneutralization assay of recombinant vesicular stomatitis virus carrying SARS-CoV-2 spike protein (rVSV-SARS2-S). a. Neutralizing antibody (nAb) titres (log10 IC50) from n = 30outpatients (116 samples; grey circles) and n = 35 hospitalized (112 samples; red circles) at 2 to 37 days post-symptom onset. c. Longitudinal nAb titres (log10 IC50) from n = 36 outpatients (85 samples) and n = 31 hospitalized (58 samples) taken from day 23 (outpatients) or day 25 (hospitalized) until day 414 postsymptom onset. c, d. The end-point titres of anti-N IgG were determined by ELISA using a recombinant SARS-CoV-2 nucleocapsid protein. Samples and time points are the same as those in A and B. a-c. The second order polynomial (quadratic) curve fitting was used to establish the days at which peak titres occurred (Ymax). b-d. Continuous decay fit is shown with the red and gray line for the corresponding patient group. Every data point represents results from two technical replicates. HRP conjugated goat anti-mouse (p/n orb345943) was used at 1:3000.

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SUMOylation of TAB2 inhibits NF-kB activation by suppressing the TRAF6/TAB2/TAK1 complex. a Dual luciferase assay analysis of the effects of TRIM60-mediated SUMOylation on TAK1/TAB2induced NF-KB activity. HEK293T cells were transiently transfected with the indicated plasmids, and the dual luciferase assay was performed. b Dual luciferase assay analysis of the effects of TAB2 mutants on the TRIM60-mediated suppression of NF-KB activity. c IP and WB analyses of the TRAF6/TAB2/TAK1 complex in control and HA-TRIM60-overexpressing RAW cells stimulated by LPS as indicated. Formation of the TRAF6/TAB2/TAK1 complex was examined in BMDMs (d) and RAW cells (e). Cells were stimulated with LPS for the indicated amounts of time, and IP and WB analyses were performed. f TRIM60 suppresses RIP1/TAB2/TAK1 signalosome formation in MEFs. IP and WB analyses of the RIP1/TAB2/TAK1 complex in MEFs. WT and TRIM60 KO MEFs were stimulated with TNF $\alpha$  as indicated, followed by IP and WB analyses. g IP and WB analyses of MAPK/NF-kB signaling activation and RIP1/TAB2/TAK1 complex formation. TAB2-deficient MEFs rescued with WT or TAB2-K329R/K562R were stimulated with TNF $\alpha$  as indicated, followed by IP and WB analyses to detect RIP1/TAB2/TAK1 complex formation, TAB2 SUMOylation, and phosphorylation of ERK and IkBa. The formation of the TRAF6/TAB2/TAK1 complex in c and d are quantified by ImageJ and shown as Supplementary Fig. 8a and b, respectively. The firefly luciferase activity levels in a and b were normalized to the Renilla luciferase activity levels and are presented as the mean ± SEM. \*\*\*P 0.001; n.s. no significance (one-way ANOVA followed by Tukey's multiple comparisons). The data are representative of three independent experiments (a-g). HRPconjugated Goat anti-mouse secondary antibody (p/n orb345943) was used.

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