

Ammonia/Ammonium Microplate Assay Kit

Cat #: orb545637 (manual)

Detection and Quantification of Ammonia/Ammonium content in Serum, Plasma, Urine, Saliva, Cell culture, Tissue extracts, Cell lysate, Other biological fluids Samples.

For research use only. Not for diagnostic or therapeutic procedures.

INTRODUCTION

Ammonia (NH_3) or its ion form ammonium (NH_4^+) is an important source of nitrogen for living systems. It is synthesized through amino acid metabolism and is toxic when present at high concentrations. In the liver, ammonia is converted to urea through the urea cycle. Elevated levels of ammonia in the blood (hyperammonemia) have been found in liver dysfunction (cirrhosis), while hypoammonemia has been associated with defects in the urea cycle enzymes (e.g. ornithine transcarbamylase). Ammonia/Ammonium Microplate Assay Kit is designed to directly measure NH_3 and NH_4^+ in a variety of samples. In the assay, ammonia reacts with hypochlorous acid, which is determined at 620nm, is directly proportional to the Ammonia/Ammonium concentration in the sample.

KIT COMPONENTS

Component	Volume	Storage
96-Well Microplate	1 plate	
Assay Buffer I	30 ml x 4	4 °C
Assay Buffer II	30 ml x 2	4 °C
Dye Reagent I	Powder x 1	4 °C
Dye Reagent II	Powder x 1	4 °C
Dye Reagent II Diluent	3 ml x 1	4 °C
Standard	Powder x 1	4 °C
Plate Adhesive Strips	3 Strips	
Technical Manual	1 Manual	

Note:

Dye Reagent I: add 7 ml distilled water to dissolve before use.

Dye Reagent II: add 3 ml Dye Reagent II Diluent into Dye Reagent II, mix before use.

Standard: add 1 ml distilled water to dissolve, then add 5 µl into 995 µl distilled water, mix, the concentration will be 1000 µmol/L.

MATERIALS REQUIRED BUT NOT PROVIDED

1. Microplate reader to read absorbance at 620 nm
2. Distilled water
3. Pipettor, multi-channel pipettor
4. Pipette tips
5. Mortar
6. Centrifuge
7. Timer

SAMPLE PREPARATION

1. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml Assay buffer I for 5×10^6 cell or bacteria, sonicate (with power 20%, sonicate 3s, interval 10s, repeat 30 times); centrifuged at 10000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and then add 0.5 ml Assay buffer II, mix.

2. For tissue samples

Weigh out 0.1 g tissue, homogenize with 1 ml Assay buffer I, centrifuged at 10000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and then add

0.5 ml Assay buffer II, mix.

3. For liquid samples

For serum, plasma, urine and other contain protein samples, add 0.1 ml sample into 0.6 ml Assay buffer I, centrifuged at 10000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and then add 0.3 ml Assay buffer II, mix.

For other sample does not contain proteins, detect directly.

ASSAY PROCEDURE

Add following reagents into the microplate:

Reagent	Sample	Standard	Blank
Sample	100 μ l	--	--
Standard	--	100 μ l	--
Distilled water	--	--	100 μ l
Dye Reagent I	70 μ l	70 μ l	70 μ l
Dye Reagent II	30 μ l	30 μ l	30 μ l
Shake and mix, put it into the oven, 37 °C for 15 minutes. Then record absorbance measured at 620 nm.			

Note:

- 1) Perform 2-fold serial dilutions of the top standards to make the standard curve.
- 2) The concentrations can vary over a wide range depending on the different samples. For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range.
- 3) Reagents must be added step by step, can not be mixed and added together.

CALCULATION

1. According to the volume of sample (contain proteins)

$$\begin{aligned}\text{NH}_3 (\mu\text{mol/ml}) &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / V_{\text{Sample}} \times 10 \\ &= 10 \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}})\end{aligned}$$

2. According to the volume of sample (does not contain proteins)

$$\begin{aligned}\text{NH}_3 (\mu\text{mol/ml}) &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / V_{\text{Sample}} \\ &= (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}})\end{aligned}$$

3. According to the cell and bacteria of sample

$$\begin{aligned}\text{NH}_3 (\mu\text{mol}/10^4) &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / (N \times V_{\text{Sample}} / V_{\text{Assay}}) \\ &= 1.5 \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / N\end{aligned}$$

4. According to the weight of sample

$$\begin{aligned}\text{NH}_3 (\mu\text{mol/g}) &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / (V_{\text{Sample}} \times W / V_{\text{Assay}}) \\ &= 1.5 \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / W\end{aligned}$$

C_{Standard} : the standard concentration, $1000 \mu\text{mol/L} = 1 \mu\text{mol/ml}$;

C_{Protein} : the protein concentration, mg/ml ;

W : the weight of sample, g ;

N : the quantity of cell or bacteria, $N \times 10^4$;

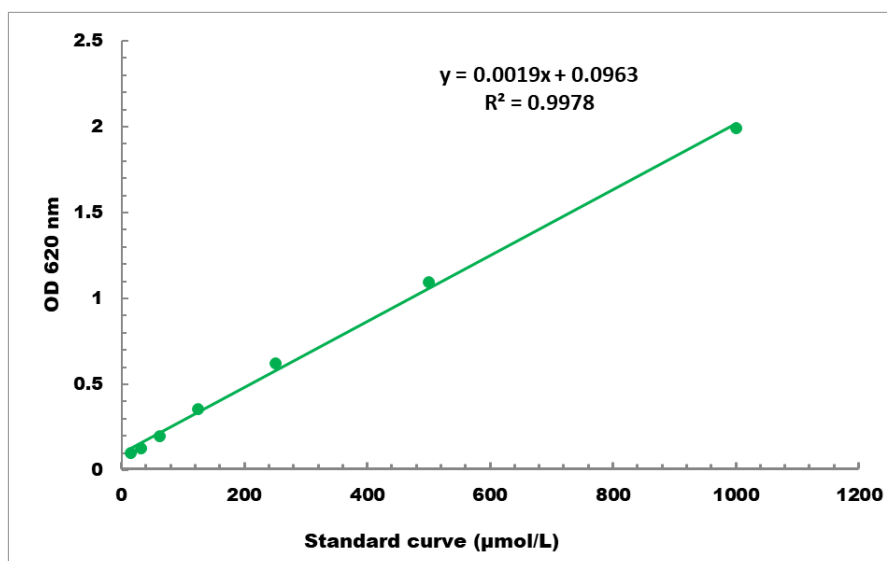
V_{Assay} : the volume of Assay buffer, 1.5 ml

V_{Standard} : the volume of standard, $100 \mu\text{l}$;

V_{Sample} : the volume of sample, $100 \mu\text{l}$.

TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 10 µmol/L - 1000 µmol/L