

Soil β-Glucosidase Assay Kit Cat#: orb1499906 (manual) Size: 100T/48S

Microassay

Product composition and storage conditions:

No.	Specifications	Storage Conditions		
orb1499906 - A	Toluene 5mL (Self- provided)	Store at 4°C		
orb1499906 - B	Powder ×2	Store at -20 ° C; Add 7.5mL distilled water to each bottle fully dissolve before use, and the remaining reagent is still stored at -20°C;		
orb1499906 - C	30mL ×1	Store at 4°C;		
orb1499906 - D	20mL ×1	Store at 4°C;		
orb1499906 - Standard	$1mL \times 1$ (5 mmol/L)	Store at 4°C.		

*Before the formal measurement, be sure to take 2-3 samples with large expected differences for predetermination.

Introduction:

Significance: B-D-Glucosidase (S- β - GC, EC3.2.1.21), also known as β -D- glucoside glucose hydrolase, gentian disaccharidase, cellodisaccharidase, cellobias, or β -G, and amygdalosidase. It belongs to cellulase and is an important component of the cellulolytic enzyme system. It can hydrolyze and bind to the terminal non-reductive β -D- glucose bond and release β -D- glucose and corresponding ligands. S- β - GC has important physiological functions in the carbohydrate metabolism of soil microorganisms.

Principle: S- β -GC catalyzes the formation of p-nitrophenol from p-nitrobenzene - β -D glucopyranoside, which is characterized by light absorption at 400 nm.

Own supplies:

Visible spectrophotometer/plate reader, table centrifuge, water bath, 30~50 mesh sieve, adjustable pipette, micro glass cuvette/96-well plate, mortar, ice, toluene and distilled water.

Sample Processing:

Fresh soil sample is naturally air-dried or 37°C oven is air-dried, passing 30~50 mesh sieve.



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Measurement steps:

- 1. Preheat the visible spectrophotometer/microplate reader for at least 30 minutes, adjust the wavelength to 400nm, set zero with distilled water.
- Preparation of standard: Take 100 μL standard solution immediately before use, add it into 400 μL reagent C to obtain 1 mmol/L standard solution (i.e. 1000 μmol/L), and then dilute with distilled water to the concentration of standard solution to be used 500, 250, 100, 50, 25 and 12.5 μmol/L.
- 3. Add the following reagents in sequence to the EP tube:

Reagent name	Measuring tube	Control tube	Standard tube	Blank tube	
Air-dried soil sample (g)	0.02	0.02			
orb1499906 – A (ul)	10	10			
Vibrate and mix well to make all soil samples wet and leave at room temperature for 15 min.					
orb1499906 – B (ul)	130				
orb1499906 – C (ul)	160	160			
Mix well, place in 37°C water bath for 1h, then boiling water bath for 5 min (cover tightly to prevent water loss), and cool down with running water.					
orb1499906 – B (ul)		130			
Fully mix, centrifuge for 10 min at 10000 g 25°C, take supernatant (Add the following reagent to the EP tube or 96well plate).					
Supernatant (ul)	70	70			
Standard (ul)			70		
Distilled water (ul)				70	
orb1499906 – D (ul)	130	130	130	130	
Mix well, leave it at room temperature for 2 minutes, measure the absorbance A at 400 nm, and calculate the $\Delta A=A$ measuring -A control, ΔA standard=A standard -A blank.					

Note: A control tube is provided for each measuring tube.

S-β-GC activity calculation:

1. Standard curve establishment:

A standard curve was established based on the concentration of the standard tube (x, μ mol/L) and the absorbance ΔA standard (y, ΔA standard). The ΔA assay (y, ΔA assay) was brought into the formula to calculate the sample concentration (x, μ mol/L) according to the standard curve.

2. Calculation of S-β-GC enzyme activity:

Definition of units: 1µmol p-nitrophenol per g soil samples per day is defined as an enzyme activity unit.

S- β -GC enzyme activity (U/g soil sample) = x × V total \div W \div T=0.36 x

T: Reaction time, 1h=1/24d; V total: Total volume of reaction system: 3×10⁻⁴L; W: Sample weight, 0.02 g.