

DXM ELISA Kit

Cat#: orb411509 (ELISA Manual)

PRINCIPLE OF THE ASSAY

This assay employs the competitive inhibition enzyme immunoassay technique.

The coupling antigen is pre-coated on the micro-well stripes. Then the Dexamethasone in the sample and the coupling antigen pre-coated on the micro-well stripes compete for the anti-Dexamethasone antibody. A substrate solution is added to the wells and the color develops in opposite to the amount of Dexamethasone in the standards or samples. The color development is stopped and the intensity of the color is measured.

DETECTION RANGE

0.1 ppb-8.1ppb

SENSITIVITY

The minimum detectable dose of the kit is typically less than 0.1ppb.

The sensitivity of this assay, or Lower Limit of Detection (LOD) was defined as the lowest concentration that could be differentiated from zero. It was determined the mean OD value of 20 replicates of the zero standard added by their three standard deviations.

CROSS-REACTION RATE

Dexamethasone	100%
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RECOVERY RATE

Muscle Tissue	80%±15%
Milk	80%±15%
Feed	80%±

LIMIT OF DETECTION

Muscle Tissue	0.2 ppb
Milk	0.5 ppb
Feed	1 ppb

PRECISION

Intra-assay Precision (Precision within an assay): CV%<10%

Three samples of known concentration were tested twenty times on one plate to assess.

Inter-assay Precision (Precision between assays): CV%<10%

Three samples of known concentration were tested in twenty assays to assess.

LIMITATIONS OF THE PROCEDURE

- The kit should not be used beyond the expiration date on the kit label.
- Do not mix or substitute reagents with those from other lots or sources.
- Any variation in operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.
- This assay is designed to eliminate interference by soluble receptors, binding proteins, and other factors present in biological samples. Until all factors have been tested in the Immunoassay, the possibility of interference cannot be excluded.

MATERIALS PROVIDED

Reagent	Quantity
Assay plate	96T
Standard	6 x 1mL
High Concentration Standard (100ppb)	1 x 1mL
HRP-conjugate	1 x 11mL
Antibody	1 x 5.5 mL
Substrate A	1 x 6mL
Substrate B	1 x 6mL
Stop Solution	1 x 6mL
Redissolving Solution (2x)	1 x 50mL
Wash Buffer(20x)	1 x 40 mL
Adhesive Strip	4
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STANDARD CONCENTRATION

Standard	S0	S1	S2	S3	S4	S5
Concentration (ppb)	0	0.1	0.3	0.9	2.7	8.1

STORAGE

Unopened kit	Store at 2 -8°C. Do not use the kit beyond the expiration date
Opened kit	May be stored for up to 1 month at 2 - 8° C.

***Provided this is within the expiration date of the kit.**

OTHER SUPPLIES REQUIRED

- Microplate reader capable of measuring absorbance at 450 nm.
- An incubator which can provide stable incubation conditions up to 25°C
- Squirt bottle, manifold dispenser, or automated microplate washer.
- Centrifuge, Vortex mixer
- Analytical balance, 2 decimal place
- Rotary evaporator or nitrogen gas
- Absorbent paper for blotting the microtiter plate.
- Single-channel micropipette (20µL-200µL, 100µL-1000µL)
- 300µL multichannel micropipette
- 100 mL and 500 mL graduated cylinders
- Deionized or distilled water.
- Pipettes and pipette tips.
- Test tubes for dilution.
- Ethyl acetate
- N-hexane
- NaOH

PRECAUTIONS

The Stop Solution provided with this kit is an acid solution. Wear eye, hand, face, and clothing protection when using this material.

Note:

- Kindly use graduated containers to prepare the reagent.
- Bring all reagents to room temperature (20-25°C) before use for 30 min.
- Only the disposable tips can be used for the experiments and the tips must be changed when used for different reagents.
- Distilled water is recommended to be used to make the preparation for reagents or samples. Contaminated water or container for reagent preparation will influence the detection result.

REAGENT PREPARATION

- **2M NaOH:** Dissolve 40.0g of NaOH to 500 mL of deionized water, shake well.
- **0.3M NaOH:** Transfer 75 mL of **2M NaOH** to 500 mL of deionized water, shake well.
- **Redissolving Solution (1x):** Take **Redissolving Solution(2x)** and deionized water (1:1), mix well. Keep it at 4°C for one month.
- **Wash Buffer (1x):** If crystals have formed in the concentrate, warm up to room temperature and mix gently until the crystals have completely dissolved. Dilute 10 mL of **Wash Buffer (20x)** into 190 mL deionized or distilled water to prepare 200 mL of **Wash Buffer (1x)**. Keep it at 4°C for one month.

Note:

1. Biorbyt is only responsible for the kit itself, but not for the samples consumed during the assay. The user should calculate the possible amount of the samples used in the whole test. Please reserve sufficient samples in advance.
2. If the samples are not indicated in the manual, a preliminary experiment to determine the validity of the kit is necessary.
3. Please predict the concentration before assaying. If values for these are not within the range of the standard curve, users must determine the optimal sample dilutions for their particular experiments.

SAMPLE COLLECTION AND STORAGE

A. Muscle Tissue

1. Weigh 2.00 ± 0.05 g of the homogenized sample, put into 50 mL centrifugal tube.
2. Add 8 mL of **Ethyl Acetate**, shake properly for 5 min.
3. Centrifuge at 4000 rpm for 10 min at room temperature
4. Transfer 4 mL of supernatant into a new centrifugal tube, add 4 mL of **2M NaOH**, shake properly for 5 min. Centrifuge at 4000 rpm for 10 min at room temperature.
5. Take 2 mL of upper layer and the sample can be dried by blowing nitrogen gas at 50-60°C.
6. Add 1 mL of **Redissolving Solution (1x)**, shake properly for 2 min.
7. Take 100 μ L of sample for further analysis.

Dilution factor of the samples: 2

B. Milk

1. Take 200 μ L of milk sample, add 0.8 mL of **Redissolving Solution (1x)**, shake well.
2. Take 100 μ L of sample for further analysis.

Dilution factor of the samples: 5

C. Feed

1. Weigh 1.00 ± 0.05 g of the homogenized feed sample.
2. Add 4 mL of **0.3M NaOH**, shake well.
3. Then add 8 mL of **Ethyl Acetate**, shake properly for 5 min.
4. Centrifuge at 4000 rpm for 10 min at room temperature.
5. Take 1 mL of upper layer and the sample can be dried by blowing nitrogen gas at 50-60°C.
6. Add 1 mL of **N-hexane**, shake well; then add 1 mL of **Redissolving Solution (1x)**, shake properly for 2 min.
7. Centrifuge at 4000 rpm for 5 min at room temperature.

8. Take 100 μ L of lower layer sample for further analysis.

Dilution factor of the samples: 8

ASSAY PROCEDURE

Bring all reagents and samples to room temperature (20~25°C) before use. Centrifuge the sample again after thawing before the assay. It is recommended that all samples and standards be assayed in duplicate.

1. Prepare all reagents and samples as directed in the previous sections.
2. Determine the number of wells to be used and put any remaining wells and the desiccant back into the pouch and seal the ziploc, store unused wells at 2-8°C.
3. Add 100 μ L of **Standard** or **Sample** per well. Then add 50 μ L of **Antibody** to each well. Incubate for 30 min at 25°C. Protect from light.
4. Aspirate each well and wash, repeating the process five times. Wash by filling each well with 250 μ L of **Wash Buffer (1x)** using a squirt bottle, multi-channel pipette, manifold dispenser, or autowasher, and let it stand for 30 seconds, complete removal of liquid at each step is essential to good performance.
5. Add 100 μ L of **HRP-conjugate** to each well, incubate for 30 min at 25°C. Protect from light.
6. Repeat the wash process as step 4.
7. Add 50 μ L of **Substrate A** and 50 μ L of **Substrate B** to each well, mix well. Incubate for 15 minutes at 25°C. Protect from light.
8. Add 50 μ L of **Stop Solution** to each well, gently tap the plate to ensure thorough mixing.
9. Determine the optical density of each well within 10 min, using a microplate reader set to 450 nm (Recommend to read the OD value at the dual-wavelength: 450/630 nm within 10 min).

Note:

1. The final experimental results will be closely related to validity of the products, operation skills of the end users and the experimental environments.
2. Samples or reagents addition: Please carefully add samples to wells and mix gently to avoid foaming. Do not touch the well wall as possible. For each step in the procedure, total dispensing time for addition of reagents or samples to the assay plate should not exceed 10 min. This will ensure equal elapsed time for each pipetting step, without interruption. Duplication of all standards and specimens, although not required, is recommended. To avoid cross-contamination, change pipette tips between additions of each standard level, between sample additions, and between reagent additions. Also, use separate reservoirs for each reagent.

3. Incubation: To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary. Do not allow wells to sit uncovered for extended periods between incubation steps. Once reagents have been added to the well strips, DO NOT let the strips DRY at any time during the assay. Incubation time and temperature must be observed.
4. Washing: The wash procedure is critical. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Solution by aspirating or decanting and remove any drop of water and fingerprint on the bottom of the plate. Insufficient washing will result in poor precision and falsely elevated absorbance reading. When using an automated plate washer, adding a 30 second soak period following the addition of wash buffer, and/or rotating the plate 180 degrees between wash steps may improve assay precision.
5. Controlling of reaction time: Observe the change of color after adding Substrates (e.g. observation once every 10 min). Substrates should change from colorless or light blue to gradations of blue. If the color is too deep, add Stop Solution in advance to avoid excessively strong reaction which will result in inaccurate absorbance reading.
6. Substrates are easily contaminated. Substrates should remain colorless or light blue until added to the plate. Please protect it from light.
7. Stop Solution should be added to the plate in the same order as the Substrates. The color developed in the wells will turn from blue to yellow upon addition of the Stop Solution. Wells that are green in color indicate that the Stop Solution has not mixed thoroughly with the Substrates.

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CALCULATION OF RESULTS

The mean values of the absorbance values obtained for the standards and the samples are divided by the absorbance value of the first standard (zero standard) and multiplied by 100%. The zero standard is thus made equal to 100% and the absorbance values are quoted in percentages.

$$\text{Absorbency value (\%)} = \frac{B}{B_0} \times 100\%$$

B -----the average absorbance value of the sample or standard

B₀-----the average absorbance value of the 0ppb standard

To draw a standard curve: Take the absorbency value of standards as y-axis, logarithmic of the concentration of the Dexamethasone standards solution (ppb) as x-axis.

The Dexamethasone concentration of each sample (ppb), which can be read from the calibration curve, is multiplied by the corresponding dilution factor of each sample followed, and the actual concentration of sample is obtained.

Note:

- Discard the substrate with any color that indicates the degeneration of this solution; when the absorbance value of standard solution 0 of less than 0.5 indicates its degeneration.
- The optimum reaction temperature is 25°C, and too high or too low will result in the changes in the absorbance value and detecting sensitivity