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## **Product Datasheet**

Normal Mouse Liver Whole Cell Lysate (orb348718)

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Normal Mouse Liver Whole Cell Lysate	
Unconjugated	
SDS-PAGE, WB	
Preservative: None. Stabilizer: 10% (v/v) Glycerol. 1X SDS-PAGE Sample Buffer (62.5 mM Tris HCl, 2% SDS, 10% Glycerol and 0.005% bromophenol blue, pH 6.8)	
Liquid (sterile filtered)	
1.0 mg/ml	
Store vial at -70° C or COLDER. For extended storage, aliquot contents to minimize freeze/thaw cycles.	
For research use only	
ready-to-use lysates are especially prepared as positive controls for separation by SDS-PAGE and subsequent western blot analysis. Lysates are prepared in denaturing buffer WITHOUT dissociating agents (i.e. no 2-mercaptoethanol or dithiothreitol has been added). Heat lysate to 95°C for 5 minutes and rapidly cool. If dissociating conditions are desired, add reducing agent prior to heating. The recommended loading volume per lane is 10-20 µl depending on the size format of your gel.	
Tissues were washed exhaustively with PBS to remove blood and other debris. A lysate was prepared by homogenizing the tissue and washing the cells in cold PBS. Washed cells were incubated at 4° C in modified RIPA buffer containing 150 mM sodium chloride, 50 mM Tris Cl, pH 7.4, 1 mM EDTA, 1.0% NP-40, 0.5% sodium deoxycholic acid and 0.1% SDS to lyse the cells. Protein integrity is ensured using a cocktail of protease inhibitors with broad specificity for the inhibition of aspartic, cysteine, and serine proteases as well as aminopeptidases (0.1 mM AEBSF HCl, 0.08 $\mu$ M Aprotinin, 5 $\mu$ M Bestatin, 1.5	
	Unconjugated SDS-PAGE, WB Preservative: None. Stabilizer: 10% (V/V) Glycerol. 1X SDS-PAGE Sample Buffer (62.5 mM Tris HCl, 2% SDS, 10% Glycerol and 0.005% bromophenol blue, pH 6.8) Liquid (sterile filtered) 1.0 mg/ml Store vial at -70° C or COLDER. For extended storage, aliquot contents to minimize freeze/thaw cycles. For research use only ready-to-use lysates are especially prepared as positive controls for separation by SDS-PAGE and subsequent western blot analysis. Lysates are prepared in denaturing buffer WITHOUT dissociating agents (i.e. no 2-mercaptoethanol or dithiothreitol has been added). Heat lysate to 95°C for 5 minutes and rapidly cool. If dissociating conditions are desired, add reducing agent prior to heating. The recommended loading volume per lane is 10-20 µl depending on the size format of your gel. Tissues were washed exhaustively with PBS to remove blood and other debris. A lysate was prepared by homogenizing the tissue and washing the cells in cold PBS. Washed cells were incubated at 4° C in modified RIPA buffer containing 150 mM sodium chloride, 50 mM Tris Cl, pH 7.4, 1 mM EDTA, 1.0% NP-40, 0.5% sodium deoxycholic acid and 0.1% SDS to lyse the cells. Protein integrity is ensured using a cocktail of protease inhibitors with broad specificity for the inhibitor of aspartic, cysteine, and serine proteases as well as aminopeptidases (0.1 mM AEBSF HCl, 0.08 µM Aprotinin, 5 µM Bestatin, 1.5

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Protein concentration was determined

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